# Blood stasis contributions to the perception of skin pigmentation

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Abstract. The chromatic characteristics of skin color arise from the interactions of light (primarily absorption and scattering) with the epidermis and the dermis. The primary light absorbers in skin are hemoglobin and melanin. Most of scattering is attributed to collagen fibers and in pigmented skin to melanosomes. Traditionally skin redness is considered to arise due to locally elevated concentrations of hemoglobin, whereas skin pigmentation is attributed to melanin. In this study we attempt to understand better the contributions of these chromophores to the perceived skin color using spectral analysis of skin color reactions induced by ultraviolet (UV) irradiation or pressure. In the first experiment 12 individuals with skin phototypes III-IV were irradiated on the back using a solar simulator with doses ranging from 0.7 to 3 MED. The skin reactions were evaluated on days 1, 7, 14, and 21 after irradiation. Evaluations included diffuse reflectance spectroscopy (DRS) and clinical assessment of the erythema and the pigment reaction. Apparent concentrations of melanin, oxy-, and deoxyhemoglobin were calculated from the absorption spectra. In the second experiment the levels of deoxy-hemoglobin of the volar forearm of ten volunteers were selectively altered by either application of a pressure cuff or by topical application of 3% H<sub>2</sub>O<sub>2</sub>. Changes in skin color appearance were documented by photography, colorimetry, and DRS. In the UV exposure experiment all reactions were dose dependent. Oxy-hemoglobin values increased to a maximum on day 1, correlating well with the clinical evaluation of erythema, and then decreased exponentially to base line. Melanin showed a significant increase on day 7 and remained relatively constant for the next 3 weeks, correlating well with the clinical evaluation of pigmentation (tanning). Deoxy-hemoglobin increased slightly on day 1 and remained elevated for the next 2 weeks. Thus, deoxy-hemoglobin correlated moderately with the clinical erythema scoring on day 1 only, while it contributes significantly to what is clinically perceived as skin tanning on days 7 and 14. Application of pressure below the diastolic level increased deoxy-hemoglobin concentration as measured by DRS. This increase corresponded to a decrease of a "pigmentation" parameter (based on the  $L^*a^*b^*$  scale) in a similar fashion that has been documented for increases in melanin concentration. Topical H<sub>2</sub>O<sub>2</sub> application reduced deoxy-hemoglobin levels as measured by DRS. This reduction coincided kinetically with a visible skin blanching. Application of pressure or H<sub>2</sub>O<sub>2</sub> did not significantly alter the levels of oxy-hemoglobin or melanin. In this report we present compelling evidence that deoxy-hemoglobin significantly contributes to the skin color appearance. Blood pooling, expressed as increased deoxy-hemoglobin, can contribute to what is visually perceived as pigmentation. Furthermore, we present that measurement of its contribution to the skin color appearance can only be accomplished with DRS. © 2004 Society of Photo-Optical Instrumentation Engineers.

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## Introduction

Human skin is structurally and optically heterogeneous. It consists of two discrete layers, the epidermis and the dermis, each with different biological structure and different optical properties arising from their individual absorbing and scattering constituents. The epidermis is a highly cellular tissue consisting of layers of keratinocytes. It is known that the primary absorber in the epidermis is melanin. It has also been shown<sup>1,2</sup> that particulate melanin is a major contributor to epidermal scattering. Melanin is synthesized in specialized organelles, the melanosomes, which are manufactured in the melanocytes that are found at the basal layer of the epidermis (Fig. 1). The melanosomes are secreted from the dendritic processes of the melanocytes and they are phagocytosed by the keratinocytes.

The dermis is essentially an acellular tissue sparsely populated by fibroblasts. It consists primarily of extracellular matrix components (collagen and elastin), blood vessels, and lymphatic vessels. Collagen microfibrils are organized into fibers and the fibers into bundles. These structures constitute the major cause of light scattering in the skin. Hemoglobin is found in the dermal blood supply and is responsible for the red appearance of skin as in the case of erythema. Skin hemoglobin is confined in networks of arterial and venous plexi that run approximately parallel to the skin surface and in small capillaries that run vertical to the skin surface and reach close to the dermal-epidermal junction in the form of capillary loops (Fig. 1). Anatomically one can distinguish a superficial arterial and venous plexus and a deeper plexus. The capillaries stem from the superficial arterial plexus and empty in the superficial venous plexus. The superficial and deeper plexi are interconnected through smaller vessels. At the deeper level arteriovenous anastomoses (shunts) provide ways for blood flow to bypass superficial skin layers, thus enabling skin ther-

mal regulation. Circulating erythrocytes in the blood vessels contain high concentrations of hemoglobin. The hemoglobin molecule has four heme groups, which can bind to and deliver oxygen molecules to the tissues. When these binding sites are unoccupied the molecule is called deoxy-hemoglobin (deoxy-Hb) and when they are occupied by oxygen molecules it is termed oxy-hemoglobin (oxy-Hb). Each form of hemoglobin has its own characteristic absorption profile.<sup>3,4</sup>

It follows from before that the visual perception of skin color is the cumulative result of contributions of various optically active molecules that are found in varying concentrations in the skin. The relative contributions of each chromophore can be evaluated quantitatively by analyzing the remittance spectra of skin tissue, keeping in mind that the calculated apparent concentrations of chromophores are averaged for a tissue volume defined by the penetration depth of the interrogating illumination.

Skin reactions are events that may be detected by the clinician's sense of vision, touch, and sometimes smell. Primarily, skin reactions such as erythema (redness), blanching (whitening), pigmentation (tanning), and induration are evaluated visually. The severity of the reaction is assessed given an index ranging typically from 0 (for no reaction) to 5 (for severe reaction). Even though the eye of the trained physician is a powerful tool such evaluations are subjective and semiquantitative. Objective quantitative evaluation of skin color reactions using noninvasive instrumentation has been used since the early decades of the 20th century.<sup>5-8</sup> Since then spectrophotometers have become smaller and simpler in use. Many researchers have used methods based on reflectance measurements that either give "erythema" and "pigmentation" indices based on simple calculations 9-12 or calculate tristimulus values ( $L^*a^*b^*$  scale) that have been adopted by

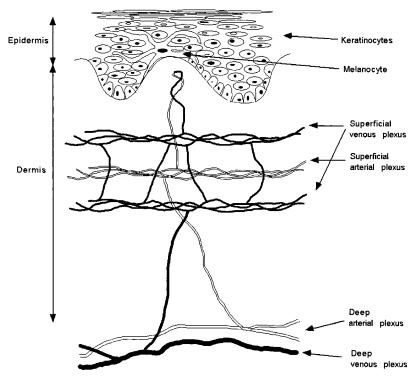


Fig. 1 Schematic diagram of skin structures.

the international committee of standards as the preferred method for color measurement.  $^{11,13-19}$  In the latter method  $L^{\ast}$  and  $b^{\ast}$ , as well as combinations of the two, have been used as pigmentation parameters and  $a^{\ast}$  as the erythema parameter. Recent studies have showed that in both methods what is clinically perceived as erythema and pigmentation does not correlate linearly with the calculated indices.  $^{20,21}$  A more accurate method is to analyze the remitted spectrum to its constituents based on diffuse reflectance spectroscopy (DRS).  $^{4,22-42}$  Apparent concentrations of melanin, oxy-Hb, and deoxy-Hb can be extracted from absorption spectra obtained by DRS, thus separating the vascular from the melanin reactions that are responsible for erythema and pigment. DRS measurements are rapid, noninvasive, objective, and quantitative. Instruments that perform DRS can be small, portable, and easy to use.

In this study we report on a number of skin reactions in which the vascular component of skin color cannot be visually separated from the melanin pigment component. Of the available methods, only spectroscopy provides the means to identify the involvement of each chromophore. In the selected examples of skin reactions presented here spectroscopically confirmed blood stasis can visually be confused with melanin pigmentation.

#### 2 Methods

#### **2.1** Clinical Studies

Twelve healthy individuals with skin phototypes III-IV participated in the study after signing an informed consent form. The study was approved by an internal review board. The source of irradiation was a 150 W UVC-filtered xenon arc solar simulator. The instrument was calibrated right before its use and the total power of the source was recorded every 3-4 h throughout the day to assure its stability and spectral quality following Colipa (The European Cosmetic, Toiletry, and Perfumery Association) guidelines. Initially the minimum solar simulator radiation (SSR) dose to induce perceptible erythema (MED) was determined on the back of each participant. Clinical erythema was evaluated 24 h after the irradiation. Following MED determination each individual was irradiated on the back with SSR doses of 0.7, 1.0, 1.5, 2.1, and 3 MED. The skin reactions were evaluated on days 1, 7, 14, and 21 after exposure. Evaluations included cross-polarized photography, DRS measurements, and clinical assessment of erythema and pigmentation by an experienced dermatologist.

In a second experiment, a pressure cuff was applied to the upper arm of ten healthy volunteers. The applied pressure was set at levels of 0, 20, 30, 40, and 60 mm Hg. Measurements were taken 5 min after each pressure level was set to allow for the vasculature to equilibrate. Changes in skin color due to vascular reactions were evaluated visually, with a chromameter, and with a DRS instrument.

In a third experiment, 2 in. diameter cotton pads soaked in 3%  $H_2O_2$  (U.S.P. topical anti-infective) in aqueous solution were applied on the volar forearm of ten healthy volunteers for 1 min. The pads were removed and the skin area was dried with fresh cotton pads. DRS measurements and visible evaluation of the treated sites were performed at 0, 5, 10, 15, and 20 min after removal of the pads.

**Table 1** Spectral characteristics of the major skin chromophores in the visible.

| Chromophore      | Absorption curve characteristic  |
|------------------|--|
| Melanin          | Monotonic increase towards short<br>wavelengths;<br>Approximates linear in the region 600–<br>750 nm |
| Oxy-hemoglobin   | Maxima at 415, 540, and 577 nm   |
| Deoxy-hemoglobin | Maxima at 430 and 555 nm   |

#### **2.2** DRS

The DRS instrument consisted of a quartz halogen light source (Ocean Optics, Boca Raton, FL), a bifurcated fiber bundle (Multimode Fiber Optics, East Hanover, NJ), an S2000 spectrometer (Ocean Optics, Boca Raton, FL), and a laptop computer (Toshiba Tecra, Irvine, CA). One leg of the fiber bundle was connected to the light source and the other to the spectrometer. Measurements were performed by placing the common end of the fiber bundle gently in contact with skin so as not to perturb the blood content. A reflectance spectrum was acquired in the range of 400–820 nm. Apparent concentrations of hemoglobin and melanin were calculated from the diffuse reflectance spectra as described elsewhere.<sup>31</sup> Briefly, the absorbance curve was calculated as the logarithm of the ratio of the diffuse reflectance from a nonirradiated site to the diffuse reflectance from an irradiated site. Pigment was evaluated from the absorbance curve as the slope of the fitted straight line over the wavelength range of 620–720 nm. Then the curve was corrected for the pigment absorption and finally, the oxy-Hb and deoxy-Hb absorption curves were fitted in the range of 550-580 nm, where they exhibit maxima (Table 1). The reproducibility of the method for calculating apparent hemoglobin concentrations was calculated as the error between measurements and was found to be better than 10%. It needs to be noted that for the collection geometry used here an underestimation of the reflectance at the long wavelengths (red/near-infrared region) compared to the shorter wavelengths (blue/green region) is anticipated. However, the measurements were always preformed relative to base line or to neighboring untreated skin and therefore such artifacts have been normalized.

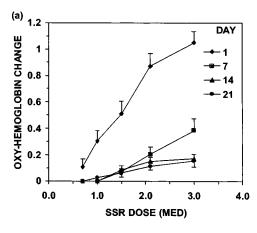
## **2.3** Data Analysis

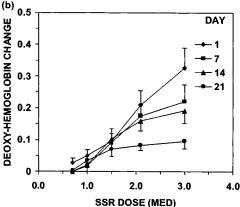
Linear regressions of the data were calculated using the least square errors algorithm. The goodness of fit is given by the correlation coefficient (R-squared). Statistical significance was calculated using the student's t-test for paired data distributions.

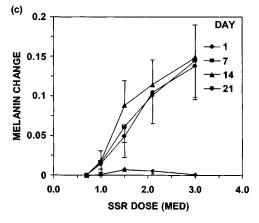
## 3 Results

# **3.1** Ultraviolet Irradiation Experiment

Ultraviolet (UV)-induced erythema is typically evaluated on day 1 after irradiation, while pigmentation is evaluated on day 7. Figure 2 shows a typical skin reaction on the back of a volunteer on days 1 and 7. The time course of changes in the concentration of oxy-Hb, deoxy-Hb, and melanin are shown

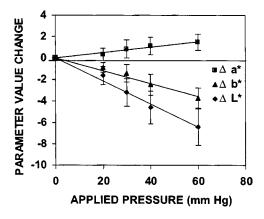






**Fig. 3** Erythema on day 1 after solar simulated exposure is accompanied by oxy- and deoxy-hemoglobin increase, whereas the observed pigment on day 7 and 14 is the combination of both melanin and deoxy-hemoglobin absorption. Shown here are the concentration changes from baseline and at different time-points of: (a) oxy-hemoglobin, (b) deoxy-hemoglobin, and (c) melanin. The data represent the average calculated values from 20 individuals who received SSR doses of 0.7, 1.0, 1.5, 2.1, and 3.0 MED on day 0. All values presented were referenced to base line (day 0).

in Fig. 3. On day 1 after irradiation the observed skin reaction was classified as clinical erythema [Fig. 2(a)]. The visual observation of erythema correlated well with a dramatic increase in oxy-Hb and deoxy-Hb, as calculated from the spectroscopic data [Figs. 3(a) and 3(b)]. The observed skin reaction on day 7 was classified clinically as pigmentation [Fig. 2(b)],

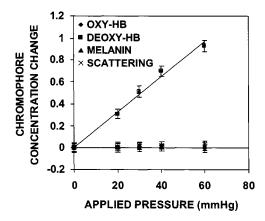


**Fig. 4** Light pressure-induced blood stasis is accompanied by skin color darkening and slight increase in the blue and the red components. Changes in the chromaticity values  $L^*a^*b^*$  of volar forearm skin were measured after application of various amounts of pressure using a pressure cuff on the upper arm. Data represent average values  $\pm$  one standard deviation acquired from ten people. Decreases in  $L^*$  represent darkening of the skin color. Decreases in  $b^*$  represent more blue and less yellow color. Increases in  $a^*$  correspond to more red and less green color. The slopes of the fitted lines are -0.107, 0.025, and -0.059 for  $L^*$ ,  $a^*$ , and  $b^*$  respectively.

which correlated with an increase in melanin [Fig. 3(c)]. However, the levels of both hemoglobins were significant on day 7 indicating that there is a strong vascular contribution to the observed reaction [Figs. 3(a) and 3(b)]. On day 14 the reaction was again classified as pigmentation and although the melanin levels remained elevated, the concentration of deoxy-Hb was still above base line [Fig. 3(b)]. On day 21 the observed pigmentation was almost exclusively due to melanin [Fig. 3(c)]. Note that although deoxy-Hb is still measurably above base line its contribution at 0.1 level is not visibly perceptible.

# 3.2 Pressure Cuff Experiment

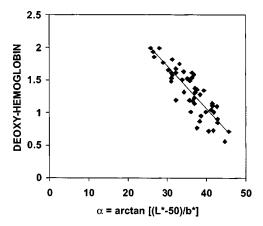
In the pressure cuff experiment increasing pressure resulted in a reduction of the values of  $L^*$  and  $b^*$  as measured by the chromameter corresponding to darker and less yellow color, respectively (Fig. 4). On the contrary the value of  $a^*$  increased corresponding to a more red appearance. Although the color changes were recorded with the chromameter it is of interest to note that visually the change in skin color was hard to observe. The reason is that the human eye works better in contrast and since the pressure was applied on the whole of the arm the color change was uniform and therefore difficult to detect. DRS analysis showed that the only chromophore that was affected by changing the applied pressure was deoxy-Hb, which increased linearly with pressure (Fig. 5). Oxy-Hb and melanin remained practically unaltered. The characteristic angle,  $\alpha = \arctan[(L^*-50)/b^*]$ , is a measure of the perceived pigmentation in such a way that when apparent pigmentation increases this parameter decreases. 13,41,43,44 In the present experiment the characteristic angle decreased with increasing pressure in all volunteers, indicating that pressureinduced increases in blood stasis can be perceived as increased pigmentation (Fig. 6).



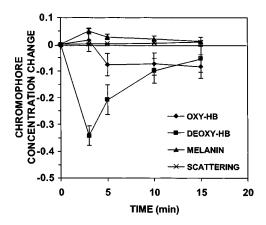
**Fig. 5** Light pressure-induced blood stasis is accompanied by a linear increase in deoxy-hemoglobin alone. Changes in the skin chromophore values of the volar forearm were measured after application of various amounts of pressure using a pressure cuff on the upper arm. Deoxy-hemoglobin increases linearly with the applied pressure. The slope of the fitted line is 0.016. Subdiastolic levels of applied pressure do not affect oxy-hemoglobin, melanin, and dermal scattering. Data represent average base line-subtracted values  $\pm$  one standard deviation acquired from ten people.

# 3.3 Hydrogen Peroxide Experiment

Application of cotton pads soaked in 3% hydrogen peroxide for 1 min induced a decrease in perceptible skin pigmentation (Fig. 7) that lasted for 10-15 min after removal of the pads. Analysis of DRS spectra showed that deoxy-Hb significantly decreased during the period of blanching (Fig. 8). Oxy-Hb decreased slightly, but within instrument variability (determined to be  $\pm\,0.1$  oxy-Hb units). Melanin and dermal scattering remained unchanged at base line levels.



**Fig. 6** The characteristic angle  $\alpha = \arctan[(L^*-50)/b^*]$ , a measure of the perception of "skin pigmentation," depends on deoxy-hemoglobin concentration in a similar fashion to its dependence on melanin. Pressure-induced changes in deoxy-hemoglobin are plotted as a function of the changes of the characteristic angle  $\alpha$ . Higher concentrations of deoxy-hemoglobin correspond to lower values of the characteristic angle, which represent perception of a more pigmented appearance. The fitted line is given by the equation y = -0.065x + 3.673.



**Fig. 8** Topical application of 3%  $H_2O_2$  reduces blood stasis as evidenced by reduction in deoxy-hemoglobin levels. Changes in skin chromophore concentrations are plotted over time. Data represent average values  $\pm$  one standard deviation acquired from ten people. Deoxy-hemoglobin values decreased with a minimum around 3 min and remain significantly lower than baseline until 15 min after  $H_2O_2$  application. Oxy-hemoglobin decreased slightly. Melanin and dermal scattering were not significantly affected.

# 4 Discussion

Melanin pigmentation is the brown-black pigment that contributes to the appearance of the skin. Melanin is produced in epidermal melanocytes in specialized vesicles called melanosomes, which are then transferred to the keratinocytes. Skin darkening is believed to be due to the distribution of melanin in the epidermis. In this study we have conducted several experiments that point out to the fact that the perceived skin darkening does not depend only on the concentration of melanin, but it is strongly affected by the concentration of deoxy-Hb in the superficial venous plexus.

We have identified a number of cases where blood pooling (stasis) and specifically the pooling of deoxy-Hb causes the appearance of the skin to get darker. As an example we determined that following solar simulated UV exposure at superthreshold levels local concentrations of deoxy-Hb confuse the assessment of melanin pigmentation at times greater than 7 days after exposure. Blood pooling contributes to the "tanned" skin appearance even 2 weeks after SSR exposure. The pooling response remains dose dependent.

Increases in blood stasis ("blood pooling") by application of a pressure cuff on the arm increases the "pigmented" appearance of the skin, as measured using a chromameter. It should be noted here that the  $L^*a^*b^*$  scale provided by the chromameter measures color as perceived by the human eye (termed also psychophotometric). Furthermore, decreases in blood stasis induced after application of hydrogen peroxide appears as pigment reduction or pigment "bleaching."

We have determined that the concentration of deoxy-Hb in the skin may be varied independently from the concentration of oxy-Hb by applying topically hydrogen peroxide, this phenomenon lasts 10–20 minutes. 44,45 It is believed that the venules contract following the application of hydrogen peroxide. Hydrogen peroxide probably interacts with the NO receptors causing the endothelial cells to contract. The reason why the arterioles are not affected may be attributed to the stronger arteriolar wall integrity that is supported by the smooth

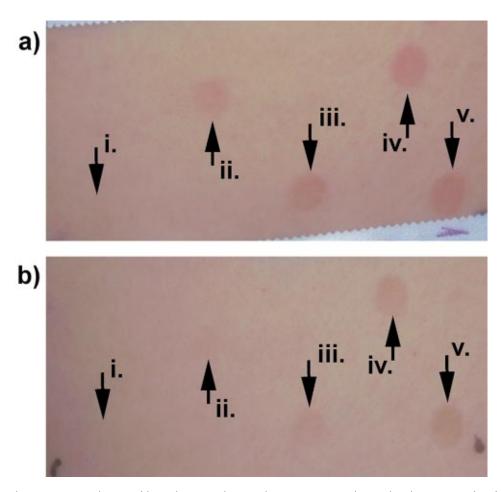
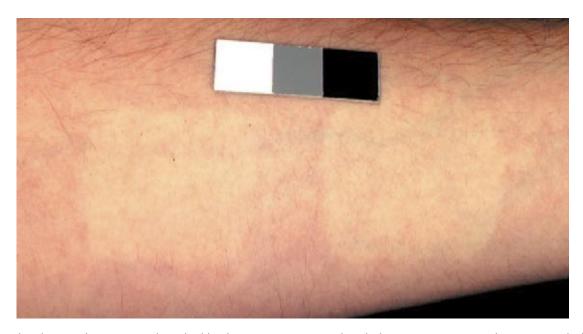
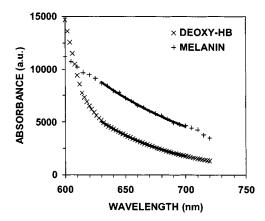


Fig. 2 Solar simulated UV exposure induces visible erythema on day 1 and pigmentation on day 7. The photos were taken from the back of an individual on (a) day 1 and (b) day 7 after irradiation. The doses from left to right were: (i) 0.7, (ii) 1.0, (iii) 1.5, (iv) 2.1, and (v) 3.0 MED. Erythema is prominent on day 1, while on day 7 the reaction was classified as pigmentation.



 $\textbf{Fig. 7} \ \, \textbf{Topical application of 3\% H}_2O_2 \ \, \textbf{results in skin blanching reaction.} \ \, \textbf{Cotton pads soaked in 3\% H}_2O_2 \ \, \textbf{aqueous solution were applied for 1 min}$ on the volar forearm. The photo was taken 2 min after removing of the soaked pads and drying the skin with fresh dry pads. Blanching with clear boundaries is evident where H<sub>2</sub>O<sub>2</sub> has been applied.



**Fig. 9** Deoxy-hemoglobin ( $\times$ ) absorbs light in a similar manner to melanin (+) in the spectral region 630–700 nm. The deoxy-hemoglobin curve was measured from a cuvette filled with lysed red blood cells under gentle nitrogen bubbling to reduce oxy-hemoglobin (see Ref. 4). The melanin curve was measured from a vitiligo patient and represents the absorbance of normally melanized skin referenced to neighboring vitiligo-involved skin (see Ref. 4). The fitted curves are given by the equations (where  $\lambda$  is the wavelength in nm): (a) melanin absorbance  $e^{15-0.009}$   $\lambda$  with  $R^2=0.9962$  and (b) deoxy-hemoglobin absorbance  $e^{18-0.015}$   $\lambda$  with  $R^2=0.9998$ .

muscle layer. An alternate mechanism would be to stimulate an enlargement of the collagen matrix or of the water holding capacity of the dermal glycosaminoglycans that surround the dermal venules. As the tissue expands it compresses the venules resulting in a partial vascular collapse. This collapse of the superficial venules results in a reduction of the average concentration of deoxy-Hb in the skin which in turn results in a lightening of the skin.

The observation that what is perceived as UV-induced pigmentation may contain a vascular component agrees with recently published reports. AC Conversely, pigmentation contributes to perceived skin redness as measured by the  $a^*$  parameter. Such observations emphasize the need for a more reliable method than the  $L^*a^*b^*$  values for quantifying erythema in the presence of pigmentation. Diffuse reflectance spectroscopy and analysis is the only available method to assess erythema and pigmentation independently through the calculation of hemoglobin and melanin concentrations. Sec. 26,31,48

The main reason why deoxy-Hb contributes to a pigmented skin appearance is that the absorption spectrum of deoxy-Hb in the 630–700 nm range is very similar to the absorption spectrum of epidermal melanin, so whatever light is transmitted has the color balance of pigment (Fig. 9). Furthermore, the size of the vessels in the superficial venous plexus is such that the transmitted radiation through these vessels is approximately 50% lower than the incident intensity and therefore they appear dark. Moreover, deoxy-Hb is a dominant pigment in normal skin contributing 30%–50% to the perceived concentration of blood and equal in its contribution to that of oxy-Hb and melanin to skin color.

We conclude therefore that the dark appearance of the skin is not only due to melanin pigment but in many cases also due to deoxy-Hb. This can be verified by pressing down on the skin surface using a glass slide, a technique termed "diascopy." This procedure compresses the vessels and drains them

temporarily making it possible to visualize the contribution of deoxy-Hb to skin color.

Because epidermal melanin and deoxy-Hb play at times equal roles in the pigment reactions of the skin we have developed instrumentation to distinguish between these two pigments. The instrument consists of a diffuse reflectance spectrometer fitted with optical fibers in order to enable measurement of any skin site. Algorithms have been developed to analyze the spectra obtained for deoxy-Hb and for melanin. In one approach we have showed that the diffuse reflectance spectra may be fitted using the diffusion approximation to account for scattering and with linear combinations of the absorption spectra of individual chromophores. <sup>40</sup> This approach while adequate does not fit the obtained spectra perfectly. The preferred approach is to fit the spectrum in sections within a narrow spectral band in order to minimize the effects of variations due to scattering in the dermis.

The results presented in this report provide strong evidence that mixed vascular and pigment reactions cannot be visually separated. Furthermore, blood stasis alone can be confused with pigmentation. In both of these situations diffuse reflectance spectroscopy provides the only means for identification of the relative involvement of each chromophore.

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### References

- M. Rajadhyaksha, M. Grossman, D. Esterowitz, R. H. Webb, and R. R. Anderson, "In vivo confocal scanning laser microscopy of human skin: melanin provides strong contrast," J. Invest. Dermatol. 104, 946–952 (1995).
- N. Kollias, R. M. Sayre, L. Zeise, and M. R. Chedekel, "Photoprotection by melanin," J. Photochem. Photobiol., B 9, 135–160 (1991).
- R. R. Anderson and J. A. Parrish, "The optics of human skin," J. Invest. Dermatol. 77, 13–19 (1981).
- N. Kollias, "The physical basis of skin color and its evaluation," Clin. Dermatol. 13, 361–367 (1995).
- L. A. Brunsting and C. Sheard, "The color of the skin as analyzed by spectrophotometric methods II. The role of pigmentation," *J. Clin. Invest.* 7, 575–592 (1929).
- L. A. Brunsting and C. Sheard, "The color of the skin as analyzed by spectrophotometric methods III. The role of superficial blood," *J. Clin. Invest.* 7, 575–592 (1929).
- E. A. Edwards and S. Q. Duntley, "The pigments and color of human skin," Am. J. Anat. 65, 1–33 (1939).
- E. A. Edwards, N. A. Finklestein, and S. Q. Duntley, "Spectrophotometry of living human skin in the ultraviolet range," *J. Invest. Dermatol.* 16, 311–321 (1950).
- P. M. Farr and B. L. Diffey, "Quantitative studies on cutaneous erythema induced by ultraviolet radiation," *Br. J. Dermatol.* 111, 673–682 (1984).
- H. Takiwaki and J. Serup, "Measurement of color parameters of psoriatic plaques by narrow-band reflectance spectrophotometry and tristimulus colorimetry," Skin Pharmacol. 7, 145–150 (1994).
- H. Takiwaki, L. Overgaard, and J. Serup, "Comparison of narrowband reflectance spectrophotometric and tristimulus colorimetric measurements of skin color," Skin Pharmacol. 7, 217–225 (1994).
- B. L. Diffey, R. J. Oliver, and P. M. Farr, "A portable instrument for quantifying erythema induced by ultraviolet radiation," *Br. J. Derma*tol. 111, 663–672 (1984).
- S. B. Park, D. H. Suh, and J. I. Youn, "A long-term time course of colorimetric evaluation of ultraviolet light-induced skin reactions," *Clin. Exp. Dermatol.* 24, 315–320 (1999).

- 14. M. J. Waring, L. Monger, D. A. Hollingsbee, G. P. Martin, and C. Marriott, "Assessment of corticosteroid-induced skin blancing: evaluation of the Minolta Chromameter CR200," Int. J. Pharm. 94, 211-222 (1993).
- 15. A. Fullerton, T. Fischer, A. Lahti, K. P. Wilhelm, H. Takiwaki, and J. Serup, "Guidelines for measurement of skin colour and erythema," Contact Dermatitis 35, 1-10 (1996).
- 16. W. Westerhof, B. A. A. M. van Hasselt, and A. Kammeijer, "Quantification of UV-induced erythema with a portable computer controlled chromameter," Photodermatol 3, 310-314 (1986).
- 17. J. Serup and T. Agner, "Colorimetric quantification of erythema-a comparison of two colorimeters with a clinical scoring scheme and laser-Doppler flowmetry," Clin. Exp. Dermatol. 15, 267–272 (1990).
- 18. J. C. Seitz and C. G. Whitmore, "Measurement of erythema and tanning responses in human skin using a tri-stimulus colorimeter," Dermatologica 177, 70-75 (1988).
- 19. P. Clarys, K. Alewaeters, R. Lambrecht, and A. O. Barel, "Skin color measurements: comparison between three instruments: the Chromameter, the DermaSpectrometer and the Mexameter," Skin Res. Technol. 6, 230-238 (2000).
- 20. J. K. Wagner, C. Jovel, H. L. Norton, E. J. Parra, and M. D. Shriver, "Comparing quantitative measurements of erythema, pigmentation and skin response using reflectometry," Pigment Cell Res. 15, 379-
- 21. H. Takiwaki, Y. Miyaoka, H. Kohno, and S. Arase, "Graphic analysis of the relationship between skin colour change and variations in the amounts of melanin and haemoglobin," Skin Res. Technol. 8, 78-83
- 22. P. Bjerring and P. H. Andersen, "Skin reflectance spectrophotometry," Photodermatol 4, 167-171 (1987).
- 23. P. H. Andersen and P. Bjerring, "Noninvasive computerized analysis of skin chromophores in vivo by reflectance spectroscopy," Photodermatol. Photoimmunol. Photomed. 7, 249-257 (1990).
- 24. P. H. Andersen and P. Bjerring, "Spectral reflectance of human skin in vivo," Photodermatol. Photoimmunol. Photomed. 7, 5–12 (1990).
- 25. P. H. Andersen, "Reflectance spectroscopic analysis of selected experimental dermatological models with emphasis on cutaneous vascular reactions," Skin Res. Technol. 3 Suppl. No. 1, 3–58 (1997).
- 26. M. G. Sowa, A. Matas, B. J. Schattka, and H. H. Mantch, "Spectroscopic assessment of cutaneous hemodynamics in the presence of high epidermal melanin concentration," Clin. Chim. Acta 317, 203-212 (1969).
- 27. N. Kollias and A. Baqer, "An experimental study of the changes in pigmentation in human skin in vivo with visible and near infrared light," Photochem. Photobiol. 39, 651-659 (1984).
- 28. N. Kollias and A. Baqer, "Spectroscopic characteristics of human melanin in vivo," J. Invest. Dermatol. 85, 38-42 (1985).
- N. Kollias and A. Baqer, "On the assessment of melanin in human skin in vivo," Photochem. Photobiol. 43, 49-54 (1986).
- 30. N. Kollias and A. H. Baqer, "Absorption mechanisms of human melanin in the visible, 400-720 nm," J. Invest. Dermatol. 89, 384-388 (1987).
- 31. N. Kollias and A. H. Baqer, "Quantitative assessment of UV-induced pigmentation and erythema," *Photodermatol* 5, 53-60 (1988).
- N. Kollias, A. Baqer, I. Sadiq, and R. M. Sayre, "In vitro and in vivo ultraviolet-induced alterations of oxy- and deoxyhemoglobin," Photochem. Photobiol. 56, 223-227 (1992).
- 33. N. Kollias, A. Baqer, and I. Sadiq, "Minimum erythema dose deter-

- mination in individuals of skin type V and VI with diffuse reflectance spectroscopy," Photodermatol. Photoimmunol. Photomed. 10, 249-254 (1994).
- 34. N. Kollias, R. Gillies, J. A. Muccini, R. K. Uyeyama, S. B. Phillips, and L. A. Drake, "A single parameter, oxygenated hemoglobin, can be used to quantify experimental irritant-induced inflammation," J. Invest. Dermatol. 104, 421-424 (1995).
- 35. N. Kollias, Y. H. Malallah, H. al-Ajmi, A. Baqer, B. E. Johnson, and S. Gonzalez, "Erythema and melanogenesis action spectra in heavily pigmented individuals as compared to fair-skinned Caucasians," Photodermatol. Photoimmunol. Photomed. 12, 183-188 (1996).
- 36. N. Kollias, R. Gillies, J. A. Muccini, S. B. Phillips, and L. A. Drake, "Oxyhemoglobin is a quantifiable measure of experimentally induced chronic tretinoin inflammation and accommodation in photodamaged skin," Skin Pharmacol. 10, 97-104 (1997).
- 37. N. Kollias and J. L. Bykowski, "Immediate pigment darkening thresholds of human skin to monochromatic (362 nm) ultraviolet A radiation are fluence rate dependent," Photodermatol. Photoimmunol. Photomed. 15, 175-178 (1999).
- 38. N. Kollias, G. N. Stamatas, and J. I. Youn, "Suppression of UVBinduced cutaneous erythema by a previous UVB exposure," Photochem. Photobiol. 74, 471-476 (2001).
- R. Gillies, W. D. Tian, R. R. Anderson, L. A. Drake, and N. Kollias, "Noninvasive monitoring of treatment related changes in psoriatic plaques using fluorescence excitation and diffuse reflectance spectroscopy," in Trends in Optics and Photonics: Biomedical Optical Spectroscopy and Diagnosis, E. M. Sevick-Muraca, J. A. Izatt, and M. N. Edinger, Eds., pp. 113-115, Optical Society of America, Washington, DC (1998).
- 40. G. Zonios, J. Bykowski, and N. Kollias, "Skin melanin, hemoglobin, and light scattering properties can be quantitatively assessed in vivo using diffuse reflectance spectroscopy," J. Invest. Dermatol. 117, 1452-1457 (2001).
- 41. J. B. Dawson, D. J. Barker, D. J. Ellis, E. Grassam, J. A. Cotterill, G. W. Fisher, and J. W. Feather, "A theoretical and experimental study of light absorption and scattering by in vivo skin," Phys. Med. Biol. 25, 695-709 (1980).
- 42. S. B. Park, C. H. Huh, Y. B. Choe, and J. I. Youn, "Time course of ultraviolet-induced skin reactions evaluated by two different reflectance spectrophotometers: DermaSpectrophotometer and Minolta spectrophotometer CM-2002," Photodermatol. Photoimmunol. Photomed. 18, 23-28 (2002).
- 43. A. Chardon, I. Cretois, and C. Hourseau, "Skin color typology and suntanning pathways," Int. J. Cosmet. Sci. 13, 191-208 (1991).
- 44. E. Tegner, "Induction of skin blanching by hydrogen peroxide," Acta Derm Venereol 74, 474-475 (1994).
- 45. D. K. Goette, "Hydrogen peroxide-induced skin blanching," Arch. Dermatol. 112, 1788-1789 (1976).
- 46. J. K. Wagner, E. J. Parra, H. L. Norton, C. Jovel, and M. D. Shriver, "Skin responses to ultraviolet radiation: effects of constitutive pigmentation, sex, and ancestry," Pigment Cell Res. 15, 385-390
- 47. S. Alaluf, D. Atkins, K. Barrett, M. Blount, N. Carter, and A. Heath, "The impact of epidermal melanin on objective measurements of human skin color," Pigment Cell Res. 15, 119-126 (2002).
- M. Ferguson-Pell and S. Hagisawa, "An empirical technique to compensate for melanin when monitoring skin microcirculation using reflectance spectrophotometry," Med. Eng. Phys. 17, 104-110 (1995).